Running Header: Incorporating traits in aquatic biomonitoring

Incorporating Traits in Aquatic Biomonitoring to Enhance Causal Diagnosis and Prediction

Joseph M. Culp,*†‡ David G. Armanini, ‡ Michael J. Dunbar, § Jessica M. Orlofske, ‡ N. LeRoy Poff, || Amina I. Pollard, # and Adam G. Yates, †† and Grant C. Hose,‡‡

†Environment Canada (NWRI/Water S&T), Fredericton, Canada
‡Canadian Rivers Institute, and Dept. Biology, Univ. New Brunswick, Fredericton, Canada
§Centre for Ecology and Hydrology, Wallingford, UK
||Colorado State University, Fort Collins, USA
#USEPA, Office of Research and Development, Washington, DC, USA
††Environment Canada, CCIW, Burlington, Canada
‡‡Macquarie University, Sydney, Australia

*To whom correspondence may be addressed: Joseph M. Culp, Environment Canada, Dept. Biology, Univ. New Brunswick, 10 Bailey Drive, Fredericton, NB, Canada, E3B 5A3. Telephone (506) 458-7458; Fax (506) 458-7597; email joseph.culp@ec.gc.ca.

ABSTRACT

The linkage of trait responses to stressor gradients promises to expand biomonitoring approaches beyond traditional taxonomic-based assessments that identify ecological effect, to providing a causal diagnosis. Traits-based information may have several advantages over taxonomic-based methods. These include providing mechanistic linkages of biotic responses to environmental condition, consistent descriptors or metrics across broad spatial scales, more seasonal stability compared with taxonomic measures, and seamless integration of traits-based analysis into assessment programs. A traits-based biomonitoring approach does not require a new biomonitoring framework because contemporary biomonitoring programs gather the basic site by species composition matrices required to link community data to the traits database. An impediment to the adoption of traits-based for biomonitoring relates to the availability, consistency, and applicability of existing trait data. For example, traits generalizations among taxa across biogeographical regions are rare and there is no consensus related to the required taxonomic resolution and methodology for traits assessment. Similarly, we must determine if traits form suites that are related to particular stressor effects, and whether there is significant variation of traits among allopatric populations. Finally, to realize the potential of traits-based approaches in biomonitoring, a concerted effort towards standardizing terminology is required with the establishment of data-interoperability infrastructures to ease the sharing and merging of broad, geographical trait information.

Key words: Traits, Biomonitoring, Bioassessment, Causal Diagnosis and Prediction

INTRODUCTION

Biomonitoring uses biota to assess the ecological condition of the environment and has traditionally incorporated the taxonomic composition of communities as an indicator of ecological change. Following this assessment process, samples are collected from potentially impacted and reference sites, identified and enumerated. Differences in taxonomic composition among samples are then compared quantitatively using various multivariate and metric approaches to infer the level of impairment (Fig 1). Here, species identity serves as a surrogate for the suite of attributes that a species possesses, with these attributes influenced by environmental conditions and evolutionary processes. An unstated assumption of this approach is that the presence of a species indicates that it possesses the traits necessary to cope with environmental conditions at a site (see Southwood 1977).

Alternatively, the traits of taxa present at a site can be used to indicate environmental conditions, and further, infer mechanisms by which the community composition is shaped. The use of traits to indicate causal mechanisms is possible because, after stressor exposure, traits distribution at the impacted site is predicted to shift relative to the reference site (Fig. 2, see Rubach et al, this issue, for linkage of traits to toxic effects). Thus, traits that impart resistance or resilience to the stressor are favored. This filtering of traits by environmental stressors may allow biomonitoring approaches to yield mechanistic understanding rather than our current ability to simply observe that ecological change has occurred. This novel approach allows improvement to the standard biomonitoring approach by linking the taxonomic data to the traits of the taxa identified. Differences in the traits of taxa in samples can then be used to generate additional metrics of the level of impairment, as well as for diagnosis of the causes of impairment (Fig 1).

In this paper we use traits as defined by McGill (2006), where traits are a measurable property of an organism, such as body size, longevity or feeding guild, usually measured in individuals and applied comparatively across species and at broad geographic scales. Our aim is to highlight how including traits within existing biomonitoring programs can improve assessment sensitivity and prediction, and provide insight into the mechanism causing that change. Specifically, our objectives are to:

- 1. Describe how traits have been previously used in biomonitoring;
- 2. Outline the benefits of traits-based approaches in biomonitoring;
- 3. Describe the challenges (opportunities and limitations) in implementing trait information into biomonitoring; and
- 4. Describe the availability, consistency, and applicability of existing trait data.

In addressing these objectives we focus on our experiences in stream biomonitoring applied to benthic macroinvertebrates because these systems have been the basis for significant conceptual development related to biomonitoring (Rosenberg & Resh 1994). However, a robust method for bioassessment should integrate multiple levels of biological organisation, so where appropriate, we provide examples and references to other biological groups and ecosystems to demonstrate the broader applicability of these ideas.

A BRIEF HISTORY OF TRAITS IN BIOMONITORING

The use of trait information in biomonitoring is not new. Attributes of the biota collected have long been used, usually *post hoc*, to explain patterns in biomonitoring metrics (e.g., the absence of filter feeding organisms following sedimentation of aquatic habitats). However, this application of traits information has not been articulated as a traits-based approach, but has been

ad hoc, and often dependent on the biological knowledge of the analyst. There is a need to formalise the use of traits in biomonitoring, which will provide guidance to all users, and add greater sensitivity and diagnostic power for biomonitoring programs (see below).

The concept of biomonitoring in rivers is attributed to Kolkwitz and Marsson (1909), who's Saprobien System assessed organic contamination in German rivers. Observations of taxa restricted to polluted or unpolluted locations led to the classification of taxa based on the empirically defined trait of pollution sensitivity, thereby providing the first trait-based assessment of stream condition. The Saprobien System *per se* is no longer widely used (but see Friedrich 1990), but the notion of organism tolerance to assess environmental condition underpins many current biomonitoring methodologies.

As the science of biomonitoring evolved, a shift in ecological thinking toward equilibrium theory and diversity saw diversity indices (e.g., Shannon and Weaver 1949, Margalef 1958, Cairns and Dickson 1971) emerging as a tool for ecosystem assessment (Hellawell 1986). More recently, indices combining the Saprobien style, 'tolerance' of taxa and the abundance and diversity of those taxa, have emerged. This began with the Trent biotic index (Woodiwiss 1960) for macroinvertebrates in the UK, with similar indices subsequently developed for broader application in the UK (e.g., BMWP score, Armitage et al. 1983), North America (e.g., Hilsenhoff's index, Hilsenhoff 1987; 1988), Australia (e.g., SIGNAL Index, Chessman 1995; 2003), and elsewhere, and for other taxonomic groups (e.g., Protozoa, Jiang 2006; Diatoms, Kelly and Whitton 1995). Like the earlier Saprobien System, these newer indices are principally based on the trait of taxa sensitivity or tolerance to particular stressors.

Growing interest in environmental assessment, combined with increasing computational capacity led to the emergence of multimetric, and multivariate predictive models for

bioassessment. The development and formalisation of multimetric approaches for fish (IBI, Karr 1981), and macroinvertebrates (RBP, Plafkin et al. 1989) encouraged the development of 'non-taxonomic' trait-based metrics, particularly those relating to functional, trophic or ecological characteristics of the biota (such as feeding guild or life history). In contrast, most multivariate models that predict a reference condition from environmental variables assess impairment through metrics derived from the taxonomic composition of assemblages (e.g., RIVPACS, Wright et al. 1993; AUSRIVAS, Simpson and Norris 2000). Although these methods remain focussed on taxonomic composition of assemblages, they provide a simple platform for the inclusion of trait information, allowing comparisons of traits between predicted and observed assemblages.

Recognizing the influence of environmental conditions on the selection and expression organism traits (Southwood 1977; Townsend and Hildrew 1994; Poff 1997) lead to the definition of a wider array of morphological, ecological and life history traits. Research by Statzner and his collegues (Doledec et al. 1994; Doledec et al. 1999; Statzner et al. 2001) has explored the relationships among this more diverse set of macroinvertebrate trait data and properties of stream systems at various spatial and temporal scales. This work provides ecological background and support for the adoption of trait information for biomonitoring, however, it has not been widely applied directly to biomonitoring applications.

The development of the SPEAR method (Liess and Von der Ohe 2005) represents a significant development in traits-based biomonitoring. The method, and its subsequent iterations (e.g., Beketov et al. 2009), incorporate various traits of macroinvertebrate taxa, such as sensitivity to toxicants, generation time, migration ability, and presence of aquatic stages to produce a metric of the proportion of 'species at risk' at a test site (Liess and Von der Ohe 2005).

This metric is compared to that derived at a reference site. The SPEAR method has the benefit of being underpinned by reasonably detailed toxicity databases (see below), which has allowed it to be applied to a range of stressors, and consequently to have stressor specificity. Although the database on the trait of toxicant sensitivity is relatively well developed (at least for some toxicants and taxa), data on life history or ecological traits is less so and represents a major challenge to the development of traits-based biomonitoring (see below).

BENEFITS OF USING TRAITS IN BIOMONITORING

The linkage of trait responses to stressor gradients promises to expand biomonitoring approaches beyond traditional taxonomic-based assessments that identify ecological effect, to causal diagnosis assessments. Research to develop an empirical framework for such diagnoses is underway, and already suggests that traits-based information may have several advantages over taxonomic-based methods. These advantages include 1) the ability to provide mechanistic linkages of biotic responses to environmental condition and improve sensitivity, 2) consistent descriptors or metrics across broad spatial scales, 3) more seasonal and interannual stability compared with taxonomic measures, 4) the ability to seamlessly integrate traits-based analysis into current assessment programs, and 5) greater utility of biomonitoring outputs in ecological risk assessment (see Van den brink et al, this issue)

Traits-based approaches can provide mechanistic linkages between biotic patterns and environmental condition, and in so doing, improve the sensitivity of bioassessments. Greater sensitivity is achieved because an assemblage's trait composition may demonstrate a substantive change before its taxonomic composition does (for example, sublethal changes; shifts in body size, age structure or reproduction that occur without or prior to a loss of taxa). A corollary is that traits-based bioassessments may be able to detect changes in ecological condition at mildly impacted sites that taxonomic-based assessments miss. This hypothesis is all the more reasonable given that traits can be more strongly related to differences in land use (Dolédec et al. 2006) and other human related disturbances than is taxonomy (Dolédec et al. 1999). Furthermore, taxonomic-based bioassessments of ecosystems exposed to different human activities often result in similar expressions of ecological condition (Carlisle and Hawkins 2008). In these cases the mechanistic link between traits and stressors may be useful for further interpretation of assessment results and provide insight regarding the causes of impairment (Fig. 1). Further refinement of trait characterization may be required, however, before traits can be linked to stressors in a causal manner (Carlisle and Hawkins 2008).

Additionally, traits-based approaches may further enhance biomonitoring by increasing the spatial applicability of reference conditions relative to taxonomic methods. This is possible because traits composition can exhibit stability across large spatial areas such as ecoregions (Charvet et al. 2000; Statzner et al. 2005). Similarly, biological traits appear to be more stable among seasons than taxonomic composition (Bêche et al. 2006), a quality that could potentially reduce biomonitoring sampling effort. Biological traits are expected to be more stable than taxonomic composition among years, although this hypothesis remains largely untested.

Finally, a traits-based biomonitoring approach does not require the formulation of a new biomonitoring framework but rather, traits-based approaches can be integrated into current assessment programs without additional data collection or sampling effort. This is because contemporary biomonitoring programs, using standard sampling and analytical protocols, gather the basic site by species composition matrices required to link community data to the traits database, and existing databases may provide the trait information needed (Fig. 1). As a result,

current study designs, sampling protocols and laboratory procedures can still be used and perhaps, because of the wider geographical consistency of traits, more widely applied. Current reference condition models can also be used as both multimetric and multivariate methods can readily incorporate traits-based information (e.g., Carlisle and Hawkins 2008). Indeed, the only addition to the framework prior to the analysis stage is the linking of the observed taxa to a database of taxa with their traits (Fig. 1), a step that is already a part of some aquatic biomonitoring programs that include tolerance or functional group indices (e.g., Plafkin et al 1989; Schmidt-Kloiber et al. 2006). Thus, because of the above characteristics, traits have the potential to be a more powerful and sensitive biomonitoring tool than taxonomic composition in all instances excepting programs aimed at conserving biodiversity.

CHALLENGES OF INCORPORATING TRAIT INFORMATION INTO

BIOMONITORING

Incorporating trait information into the existing biomonitoring framework offers several advantages as discussed, but broad application of this approach is limited by several challenges (see also Van den Brink et al, this issue). These challenges reflect the character of trait data, including taxonomic and trait resolution, trait measurement, population variation of traits and correlation among traits. In addition, we have little knowledge of how combinations of traits may link in suites related to particular stressor effects.

Relationship of taxonomic and trait resolution

Biomonitoring programs commonly use family (e.g., Hilsenhoff 1988, Metzeling and Miller 2001, Reynoldson et al. 2001) or genus level taxonomic resolution (e.g., Carter and Resh 2001, Jones 2008), and by extension, traits-based approaches should match this resolution.

However, some argue that species level data may better detect ecological patterns and perturbations (Carter and Resh 2001, Lenat and Resh 2001, Jones 2008). Thus, the decision to include lower taxonomic identification and more detailed trait information in biomonitoring presents a particularly difficult decision for practitioners. For example, increased effort and financial resources are often required to improve taxonomic resolution in biomonitoring programs (Bailey et al. 2001, Jones 2008), especially in areas where there is limited regional taxonomic knowledge. Even if species level identification can be reasonably obtained, the corresponding basic biological information from which to derive traits data is often lacking, especially for some types of traits (e.g., physiological). This issue may not be a large concern for biomonitoring given that Gayraud et al. (2003) concluded that genus, and perhaps family, identifications for invertebrates may be adequate for traits-based approaches in river biomonitoring. It remains to be shown whether this trend holds for other taxonomic groups and aquatic ecosystems. Nevertheless, we anticipate that increased traits resolution will improve the ability to detect a response to an environmental driver up to a threshold where higher resolution results in reduced ability to detect ecological pattern. A related decision is whether to include or omit rare-taxa traits. In this case, the solution will likely mirror the well-developed arguments for the use of rare species in bioassessment (e.g., Cao et al. 1998, Marchant. 1999, Marchant 2002). Parenthetically, this decision may be less critical for traits-based analyses if traits composition is more temporally and spatially stable than taxonomy-based analyses (Charvet et al. 2000; Statzner et al. 2005).

Trait representation and measurement

Biomonitoring programs may not only differ with respect to taxonomic resolution but also in the number of taxonomic groups they consider (Carter and Resh 2001). For traits-based approaches to be effectively incorporated into monitoring programs, the effect of taxonomic bias toward a subset of taxa, such as known sensitive (e.g., Ephemeroptera, Plecoptera and Trichoptera) or easily identifiable taxa, must be understood. The use of a narrow range of taxa for biomonitoring could also limit the suite of traits available for identifying the type and level of environmental impact. It is important then that biomonitoring programs incorporate adequate taxonomic diversity in order to provide the trait diversity to make a traits-based approach effective. However, only a small range of traits may be needed for targeted monitoring.

Biomonitoring practitioners must also determine the way in which traits are quantified, e.g., on a categorical or continuous scale, with some traits being naturally categorical (e.g., foraging mode) and others continuous but coded as categorical (e.g., body size). Given the coarse nature of biomonitoring data, categorical trait data may be adequate for environmental assessment (e.g., see Dolédec et al. 1999, Bonada et al. 2006). A particular trait can often be measured in a number of ways, especially across taxonomic groups, and it is important that particular traits are measured consistently. For example, several conventions can be applied to the determining the appropriate means of quantifying particular traits, we anticipate an improvement in causal diagnostic power of traits-based assessment. Finally, a detailed definition of each trait and its related modalities must be provided with metadata detailing how the information was classified and measured so trait data can be consistently added for new taxa (Baird et al. this issue).

Trait correlations and combinations

The range of traits that can be selected for inclusion in a biomonitoring program depend on the type and amount of trait information available (see above). However, in the selection of

particular traits, it is important to recognize that many traits are not independent, i.e., they are correlated to each other (Resh et al. 1994, Poff et al. 2006). For example, long adult lifespan is correlated with semi-voltinism in lotic insects and flow preferences are correlated to several traits including body shape, armouring and attachment (see Poff et al. 2006). Trait correlations often reflect phylogenetic relationships between taxa, representing their shared evolutionary history (Blomberg et al. 2003, Buchwalter et al. 2008). Trait correlations can complicate the nature of environment-trait relationships when strong environmental selection for one trait causes other, linked traits to also respond (Poff et al. 2006).

Correlated traits may confuse inference in the trait-stressor relationship of interest to the biomonitoring program, and this can be magnified in multi-stressor conditions. We need better information on the relative sensitivity of individual and correlated traits to different stressors. In some cases, correlated traits may simply be redundant and not add to the overall information gained from using only selected traits (Poff et al. 2006). Conversely, recognition of phylogenetically correlated traits may provide a way to derive traits for understudied taxa and thus allow identification of groups of taxa that respond in a similar way to a particular stressor (Buchwalter et al. 2008). Information on trait correlations is currently available for some datasets, (lotic insects, Poff et al. 2006), but for other taxonomic groups and habitat types this information may not yet be available.

The inter-correlation among traits (with phylogenetic signature) means there are a limited number of possible trait combinations (or syndromes; Poff et al. 2006, Horrigan and Baird 2008) that actually exist in nature. This raises the possibility that the suite of traits possessed by an organism could be used to examine response to environmental stressors as an alternative to traits examined individually, depending on the stressor and environmental management goals (i.e. life

history strategies; Verberk et al. 2008a, b). In this way, the multivariate character of the trait response may also be matched by the multivariate nature of the environmental stressors.

Population variation in traits

Traits currently available and used in a biomonitoring context are frequently treated as static values, in that they do not allow for potentially important variation displayed by a population (Brown 1995, Jones et al. 2009). The variability/range of trait values within a population at a site may differ from that of populations of the same organism at different locations, possibly confounding the inferences that can be made from assigning all of a particular taxon to a fixed set of traits. Population variation may be the result of phenotypic plasticity, a genotypic response to the environment, or ontogeny (Sagnes et al 2008, Petchey et al. 2004). A clear example may be if populations are asynchronous, leading to differences in developmental stages in populations at different locations. The various sources of population variation may act disproportionately on selected traits which may result in some traits possessing more variation than others (see Blanck and Lamouroux 2007). The variation of traits within a population is an important evolutionary mechanism to allow organisms to cope with environmental change. As a result, population variation may occur in widely distributed species as a result of geographical shifts in ecological preferences (Buffagni et al., 2009) and as well for invasive species.

Ontogeny is an important source of population variation, especially for some invertebrate groups that possess different developmental pathways. Larval stages may differ in their life history characteristics and can even change from one larval stage to another (Sagnes et al. 2008). In freshwater biomonitoring, which typically uses insects and other invertebrates as metrics, traits are usually applied on the basis of the mature larval stages because taxonomic knowledge is in most cases also restricted to identification of mature larvae (see above; Merritt et al. 2008).

Some traits show clear ontogenetic change (e.g., predaceous insects are often detritivores as early instars), and failure to capture this detail can potentially limit trait applications, particularly if early life stages are disproportionately sensitive to a particular environmental stressor. However, this could be considered a limitation of taxonomy as well, since all organisms with the same taxonomic identity are generally assumed to possess the same characteristics, sometimes coarsely designated into adult versus immature categories, for example adult and larval stages of aquatic Coleoptera. Similarly, population variation could arise from sexual dimorphism and species showing multiple trait states depending on life cycle stages (e.g., sexual and asexual reproduction).

A better understanding of the sources and extent of trait variation as well as its importance for defining sensitivity to environmental stressors should lead to improved traitbased inferences. In some cases, however, variation in particular traits may even be useful or diagnostic of particular environmental stressors (Hodkinson and Jackson 2005). For example, morphological variation and certain life history characteristics have been used to detect stressors such as chemical contamination (Lenat et al. 1993) and thermal shifts (Hogg and Williams 1996).

Trait-environmental linkage

Few traits were initially described with the intention of representing a clear mechanistic linkage or response to environmental stressors (Poff et al. 2006). In fact, some traits exist simply as a result of the types of information available for particular taxa (Statzner et al. 2004). Yet, a clear mechanistic linkage between environmental gradient and biological traits has been demonstrated for the primary biological elements used for biomonitoring: benthic invertebrates

(e.g., Pollard and Yuan 2009), fish (e.g., Lamoroux et al. 2002), macrophytes (e.g., Thiebaut et al. 2002) and phytoplankton (e.g., Litchman et al. 2007).

Where a mechanistic linkage between traits and environmental stressors can be established the relationship can often be complex. Some traits might respond to multiple features of the environment. For instance, several authors have provided mechanistic explanation of morphological traits in stream invertebrates as an adaptation to flow (e.g., Hynes 1970; Dolédec *et al.* 2007). However, Horrigan and Baird (2008) demonstrated that in a multi-stressor environment, selected trait modalities were influenced exclusively by changes in flow conditions and were not responsive to thermal and oxygen stress while other traits were simultaneously responsive to the multiple stressors, and consequently had reduced diagnostic power. Similarly, life cycle strategies can provide an useful indicator of community condition, but their diagnostic power is limited because many taxa alter their life cycle in response to a multitude of stressing conditions (e.g., Füreder et al. 2007; Diaz et al. 2008, Richards et al. 1997, Doledec et al. 2006). Identifying stressor-specific traits (i.e., trait suites) based on the understanding of the causal relationship between trait occurrence and stressor level should be the focus of next generation traits developments.

AVAILABILITY, CONSISTENCY AND APPLICABILITY OF TRAITS DATA

Traits-based approaches require the association of taxa and their traits (Fig. 1), which usually requires the development of new databases or compilation of existing ones. Traits information is available for a variety of traits and taxonomic groups through published and online data, or it can be obtained from publishing authors. Table 2 provides a range of examples that illustrate the variety of available databases, traits and approaches, and the variability of geographical and taxonomic coverage. We restrict our reporting to: (1) trait databases that minimally apply to ecoregions, with the caveat that many other databases can be found for smaller geographical areas; and (2) trait databases which are of potential use in biomonitoring.

Consistency of trait description and classification is critical if the full potential of traits-based approaches is to be realised. Such consistency begins with a conceptual framework of trait definitions that are valid across different taxonomic groups and ecosystems. Unfortunately, such frameworks are still evolving for individual taxonomic groups, and traits generalizations among taxa across biogeographical regions are rare except for a limited number of traits (e.g., body size, basal metabolic rate, life duration). For biomonitoring purposes, consistency must be achieved for the taxonomic group and resolution at which the assessment is undertaken. Here, consistency refers to the (1) applicability of trait modalities to all taxa, for example some physiological traits may not be compatible across higher taxonomic groups with very different physiologies, and (2) adequate knowledge of traits differences related to taxonomic resolution (e.g., trait states of insect families and genera similar and well described).

A first step towards building such a global trait description for species is the use of a standard nomenclature. Although there are no established requirements and protocol for a nomenclature database source, a taxonomic basis is essential. Future database development will likely include molecular markers as these may be a useful foundation from which to reference existing traits information (see Baird et al, this issue). Similarly, there are no specific requirements to create a comprehensive trait and environmental database as advocated by Statzner et al. (2007), but efforts should be oriented towards developing data inter-operability standards (see Baird et al, this issue).

Several studies have grouped traits into broad categories such as life history, mobility, morphology and ecology (e.g., Lamouroux et al. 2002, Poff et al. 2006; Litchman and Klausmeier 2008; Stevenson et al., 2008), but a common categorization and vocabulary is lacking. A point in case is the broad list of traits categorized as "ecological" by several authors. For example, the set of traits used by Poff et al. (2006) includes four trait categories: life history, mobility, morphology and ecology. This last category includes traits for both habitat preference and behaviour (e.g., locomotion types). Carlisle and Hawkins (2008) similarly use the category of 'ecological traits' to include the functional attributes as defined by Poff et al. (2006), and stressor tolerance values (i.e., requirements for specific environmental parameters). The use of ecological requirement for environmental conditions is considered a trait by several authors (e.g., Poff et al. 2006; Stevenson et al. 2008; Buffagni et al., 2009), despite these preferences potentially being driven by multiple biological traits. In order for traits-based approaches to develop more fully, a concerted effort towards standardizing terminology and a theoretical framework is urgently required. Such developments are fundamental to producing a shared ontology and establishing broad geographical trait databases.

Finally, a number of analytical methods have been used in the study of traits. These may be for the definition of the traits themselves, exploration of the relationships amongst traits, the definition or calibration of trait-environment relationships, and their subsequent validation. Statistical methods used include correlation (Horrigan and Baird 2008), regression (Cowlishaw et al. 2009), ordination (Dray and Legendre 2008), and cluster analysis (Poff et al. 2006). Unfortunately, univariate correlations do not result in the direct production of a model, and are unable to disentangle multiple stressor-trait relationships, especially where traits interact. Consequently, they may have limited applicability in establishing linkages between traits and

environmental conditions. Regression and multivariate (ordination, clustering) approaches have been widely used in traits analysis, with a key difference being that regression models usually consider traits as explanatory variables, while the multivariate models treat traits as response variables. This distinction can be important when turning relationships into indices and management objectives.

CONCLUSIONS AND SUMMARY

Trait information has long been used in biomonitoring programs, but its full potential is yet to be realized. Traits-based approaches have the potential to expand the outputs of biomonitoring programs from the identification of impact to include a causal diagnosis. Importantly, this can be achieved with little change to existing monitoring programs, except the linking of the observed taxa to a database of taxa with their traits. Traits-based approaches have a number of desirable attributes, such as greater stability of assessment outcomes and broader applicability of reference conditions, which are likely to broaden their range of application and increase their sensitivity. Indeed, we have outlined many of the benefits that traits-based biomonitoring can provide such as the potential to develop consistent descriptors or metrics across broad spatial scales. However, there a number of challenges to be met for the full potential of traits-based approaches to be realized, central to which, is the establishment of effective and accessible trait databases (see Baird et al, this issue). While some databases already exist, their remains a paucity of traits data for many taxonomic groups and many types of traits, which is a clear and immediate research need. A concerted effort towards standardizing terminology is required with the establishment of data-interoperability infrastructures to ease the sharing and merging of broad, geographical trait information. Research effort also need to determine whether

generalizations can be made among taxa and across biogeographical regions, and on producing a consensus on the required taxonomic resolution and methodology for traits assessment. Clearly, there have been already a number of promising developments in traits-based biomonitoring approaches. It is our hope that the discussion provided here will be the stimulus for many more.

Acknowledgements— We are grateful to the organizers of the TERA workshop for providing opportunity for this collaboration and the development of these ideas. In particular, we thank Environment Canada, Syngenta, Wageningen UR/ Alterra, and CEFIC for providing funds to support the TERA workshop. We also thank our colleagues at that workshop for their discussions and input; you are all stars (*sensu* Hose unpublished).

REFERENCES

- Armitage PD, Moss D, Wright JF, Furse MT. 1983. The performance of a new biological water quality system based on macroinvertebrates over a wide range of unpolluted running-water sites. *Water Res* 17:333-347.
- Bailey RC, Norris RH, Reynoldson TB. 2001. Taxonomic resolution of benthic macroinvertebrate communities in bioassessments. *J N Am Benthol Soc* 20:280-286.
- Baird DJ, Baker CJO, Brua R, Hajibabaei M, McNicol K, Pascoe TJ, de Zwart D. 2010.
 Developing a knowledge infrastructure for traits-based ecological risk assessment. *Integr Environ Assess Manag* this issue.
- Bêche LA, McElravy EP, Resh VH. 2006. Long-term seasonal variation in the biological traits of benthic-macroinvertebrates in two Mediterranean-climate streams in California, USA. *Freshwater Biol* 51:56-75.
- Beketov M, Foit K, Schäfer R, Schriever C, Sacchi A, Capri E, Biggs J, Wells C, Liess M. 2009. SPEAR indicates pesticide effects in streams - comparative use of species- and familylevel biomonitoring data. *Environ Pollut* 157:1841-1848.
- Blanck A, Lamouroux N. 2007. Large-scale intraspecific variation in life-history traits of European freshwater fish. *J Biogeogr* 34:862-875.
- Blomberg SP, Garland T, Ives A. 2003. Testing for phylogenetic signal in comparative data: behavioral traits are more labile. *Evolution* 54:717-745.
- Bonada N, Prat N, Resh VH, Statzner B. 2006. Developments in aquatic insect biomonitoring: a comparative analysis of recent approaches. *Annu Rev Entomol* 51:495-523.

- Bowman MF, Bailey RC. 1997. Does taxonomic resolution affect the multivariate description of the structure of freshwater benthic macroinvertebrate communities? *Can J Fish Aquat Sci* 54:1802-1807.
- Brooks JL, Dodson SI. 1965. Predation, body size, and composition of plankton. *Science* 150:28-35.

Brown JH. 1995. Macroecology. Chicago: USA. University Of Chicago Press. 269 p.

- Buchwalter DB, Cain DJ, Martin CA, Xie L, Luoma, SN, Garland T. 2008. Aquatic insect ecophysiological traits reveal phylogenetically based differences in dissolved cadmium susceptibility. *Proc Nat Acad Sci USA* 105:8321-8326.
- Buffagni A, Cazzola M, Alba-Tercedor J, López Rodríguez MJ, Armanini DG. 2009. Vol. 3.
 Ephemeroptera. In: Schmidt-Kloiber A, Hering D, editors. Distribution and Ecological
 Preferences of European Freshwater Organisms, 1st ed. Sofia/Moscow, Bulgaria/Russia:
 Pensoft Publishers. p 1-256.
- Cairns J Jr, Dickson KL. 1971. A simple method for the biological assessment of the effects of waste discharges on aquatic bottom dwelling organisms. J Wat Pollut Control Fed 43:755-772.
- Cao Y, Williams DD, Williams NE. 1998. How important are rare species in aquatic community ecology and bioassessment? *Limnol Oceanogr* 43:1403-1409.
- Carlisle DM, Hawkins. CP. 2008. Land use and the structure of western US stream invertebrate assemblages: predictive models and ecological traits. *J N Am Benthol Soc* 27:986–999.
- Carter JL, Resh VH. 2001. After site selection and before data analysis: sampling, sorting, and laboratory procedures used in stream benthic macroinvertebrate monitoring programs by USA state agencies. *J N Am Benthol Soc* 20:658-682.

- Charvet S, Statzner B, Usseglio-Polatera P, Dumont B. 2000. Traits of benthic macroinvertebrates in semi-natural French streams: an initial application to biomonitoring in Europe. *Freshwater Biol* 43:277-296.
- Chase JM, Leibold MA. 2003. Ecological Niches. Chicago (IL), USA: University of Chicago Press. 216 p.
- Chessman BC. 1995. Rapid assessment of rivers using macroinvertebrates: A procedure based on habitat-specific sampling, family level identification and a biotic index. *Aust J Ecol* 20:122-129.
- Chessman B. 2003. SIGNAL 2 A Scoring System for Macro-invertebrate ('Water Bugs') in Australian Rivers. Monitoring River Heath Initiative Technical Report no. 31,
 Commonwealth of Australia, Department of the Environment and Heritage, Canberra. ISBN 0642548978
- Chessman B, Royal MJ. 2004. Bioassessment without reference sites: use of environmental filters to predict natural assemblages of river macroinvertebrates. *J N Am Benthol Soc* 23:599-615.
- Chevenet F, Dolédec S, Chessel D. 1994. A fuzzy coding approach for the analysis of long-term ecological data. *Freshwater Biol* 31:295-309.
- Cowlishaw G, Pettifor RA, Isaac NJB. 2009. High variability in patterns of population decline: the importance of local processes in species extinctions. *Proc R Soc London B* 276:63-69.
- Diaz AM, Alonso MLS, Gutierrez MSV. 2008. Biological traits of stream macroinvertebrates from a semi-arid catchment: patterns along complex environmental gradients. *Freshwater Biol* 53:1–21.

- Dolédec S, Statzner B. 1994. Theoretical habitat templets, species traits, and species richness:
 548 plant and animal species in the Upper Rhône River and its floodplain. *Freshwater Biol* 31: 523-538.
- Dolédec S, Statzner B, Bournard M. 1999. Species traits for future biomonitoring across ecoregions: patterns along a human-impacted river. *Freshwater Biol* 42:737-758.
- Dolédec S, Lamouroux N, Fuchs U, Mérigoux S. 2007. Modelling the hydraulic preferences of benthic macroinvertebrates in small European streams. *Freshw Biol* 52:145–164.
- Dolédec S, Phillips N, Scarsbrook MR, Riley RH, Townsend CR. 2006. A comparison of structural and functional approaches to determining land-use effects on grassland stream invertebrate communities. *J N Am Benthol Soc* 25:44-60.
- Dray S, Legendre P. 2008. Testing the species traits–environment relationships: the fourth-corner problem revisited. *Ecology* 89:3400-3412.
- Euro-limpacs Consortium. 2008. Freshwaterecology.info The taxa and autecology database for freshwater organisms. Available from www.freshwaterecology.info (3.2 08/2008).
- Friedrich G. 1990. Eine Revision des Saprobiensystems. Z. Wasser-, Abwasser-Forsch. 23, S. 141-152.
- Füreder L. 2007. Life at the Edge: Habitat Condition and Bottom Fauna of Alpine Running Waters. *Internat Rev Hydrobiol* 92:491–513.
- García-Barros E. 2008. Body size, egg size, and their interspecific relationships with ecological and life history traits in butterflies (Lepidoptera: Papilionoidea, Hesperioidea). *Biol J Linn Soc* 70:251-284
- Goldstein RM, Meador MR. 2005. Multilevel assessment of fish species traits to evaluate habitat degradation in streams of the Upper Midwest. *N Am J Fish Manage*. 25:180-194.

- Gordon ND, McMahon TA, Finlayson BL, Gippel CJ, Nathan RJ. 2004. Stream Hydrology: An Introduction for Ecologists. England: John Wiley and Sons, Ltd. 429p.
- Green RH. 1979. Sampling Design and Statistical Methods for Environmental Biologists. New York, USA: John Wiley and Sons.
- Hairston NG, Walton WE. 1986. Rapid evolution of a life-history trait. *Proc Nat Acad Sci USA* 83:4831-4833.
- Hardy PB, Sparks TH, Isaac NJB, Dennis RLH. 2007. Specialism for larval and adult consumer resources among British butterflies: Implications for conservation. *Biol Conservat* 138:440-452
- Hellawell J. 1986. Biological Indicators of Freshwater Pollution and Environmental Management. London, Elsevier.
- Henry CP, Amoros C, Bornette G. 1996. Species traits and recolonization processes after flood disturbances in riverine macrophytes. *Vegetatio* 122:13-27.
- Hilsenhoff WL. 1987. An improved biotic index of organic stream pollution. *Great Lakes Entomol* 20:31-39.
- Hilsenhoff WL. 1988. Rapid field assessment of organic pollution with a family-level biotic index. *J N Am Benthol Soc* 7:65-78.
- Hodkinson ID, Jackson JK. 2005. Terrestrial and aquatic invertebrates as bioindicators for environmental monitoring, with particular reference to mountain ecosystems. *Environ. Manage* 35:649-666.
- Hogg ID, Williams DD. 1996. Response of stream invertebrates to a global-warming thermal regime: an ecosystem-level manipulation. *Ecology* 77:395-407.

- Horrigan N, Baird DJ. 2008. Trait patterns of aquatic insects across gradients of flow- related factors: a multivariate analysis of Canadian national data. *Can J Fish Aquat Sci* 65:670-680
- Hynes HBN. 1970. The ecology of running waters. Toronto (ON), Canada: Toronto University Press. 555 p.
- Jiang J-G. 2006. Development of a new biotic index to assess freshwater pollution. *Environ Pollut* 139:306-317
- Jones CF. 2008. Taxonomic sufficiency: The influence of taxonomic resolution on freshwater bioassessments using benthic macroinvertebrates. *Environ Rev* 16:45-69.
- Jones KE, Bielby J, Cardillo M, Fritz SA, O'Dell J, Orme CDL, Safi K, Sechrest W, Boakes EH,
 Carbone C, Connolly C, Cutts MJ, Foster JK, Grenyer R, Habib M, Plaster CA, Price SA,
 Rigby EA, Rist J, Teacher A, Bininda-Emonds ORP, Gittleman JL, Mace GM, Purvis A.
 2009. PanTHERIA: a species-level database of life history, ecology, and geography of
 extant and recently extinct mammals. *Ecology* 90:2648-2648.
- Kelly MG, Whitton BA. 1995. The Trophic Diatom Index: a new index for monitoring eutrophication in rivers. J *Appl Phycol* 7:433-444.
- Kolkwitz R, Marsson K. 1909. Ökologie der tierischen Saprobien. Beiträge zur Lehre von des biologischen Gewasserbeurteilung. Int Rev der gesamten Hydrobiol Hydrog, 2:126-152.
- Lamouroux N, Poff NL, Angermeier PL. 2002. Convergence of stream fish community traits in France and Virginia (USA) streams along hydraulic and geomorphic gradients. *Ecology* 83:1792–1807.
- Lenat DR. 1993. Using mentum deformities of chironomus larvae to evaluate the effects of toxicity and organic loading in streams. *J N Am Benthol Soc* 12:265-269.

- Lenat DR, Resh VH. 2001. Taxonomy and stream ecology- The benefits of genus- and specieslevel identifications. *J N Am Benthol Soc* 20:287-298.
- Liess M, Von der Ohe PC. 2005 Analyzing effects of pesticides on invertebrate communities in streams. *Environ Toxicol Chem* 24:954–65.
- Litchman E, Klausmeier CA. 2008. Trait-based community ecology of phytoplankton. *Annu Rev Ecol Evol Syst* 39:615-639.
- Litchman E, Klausmeier CA, Schofield OM, Falkowski PG. 2007. The role of functional traits and trade-offs in structuring phytoplankton communities: scaling from cellular to ecosystem level. *Ecol Lett* 10:1170-1181.
- Marchant R. 1999. How important are rare species in aquatic community ecology and bioassessment? A comment on the conclusions of Cao et al. *Limnol Oceanogr* 44:1840-1841.
- Marchant R. 2002. Do rare species have any place in multivariate analysis for bioassessment? J. *N Am Benthol Soc* 21:311-313.
- Margalef JL. 1958. Information theory in ecology. General Systems 3:36-71.
- McGill BJ, Enquist B, Weiher E, Westoby M. 2006. Rebuilding ecology from functional traits. *Trends Ecol Evol* 21:178-185.
- Merritt RW, Cummins KW, Berg MB. 2008 An introduction to the aquatic insects of North America. Dubuque, Iowa, USA: Kendall Hunt Pub Co. 1158 p.
- Merritt DM, Scott ML, Poff L, Auble GT, Lytle DA. 2009. Theory, methods and tools for determining environmental flows for riparian vegetation: riparian vegetation-flow response guilds. *Freshwater Biol* doi:10.1111/j.1365-2427.2009.02206.x

- Metzeling L, Miller J. 2001. Evaluation of the sample size used for the rapid bioassessment of rivers using macroinvertebrates. *Hydrobiologia* 444:159-170.
- Murray BR, Hose GC. 2005a. The inter-specific range size-body size relationship in Australian frogs. *Global Ecol Biogeogr* 14:339-345.
- Murray BR, Hose GC. 2005b. Life-history and ecological correlates of decline and extinction in the endemic Australian frog fauna. *Austral Ecol* 30:564-571.
- Petchey OL, Downing AL, Mittelbach GG, Persson L, Steiner CF, Warren PH, Woodward, G.
 2004. Species loss and the structure and functioning of multitrophic aquatic systems. *Oikos* 104:467-478.
- Plafkin JL, Barbour MT, Porter KD, Gross SK, Hughes RM. 1989. Rapid bioassessment protocols for use in streams and rivers: Benthic Macroinvertebrates and Fish. EPA/444/4-89-001 U.S. Environmental Protection Agency, Washington.
- Poff NL, Allan JD. 1995. Functional organization of stream fish assemblages in relation to hydrologic variability. *Ecology* 76:606-627.
- Poff NL. 1997. Landscape filters and species traits: towards mechanistic understanding and prediction in stream ecology. *J N Am Benthol Soc* 16:391-409.
- Poff NL, Olden JD, Vieira NKM, Finn DS, Simmons MP, Kondratieff BC. 2006. Functional trait niches of North American lotic insects: trait-based ecological applications in light of phylogenetic relationships. J N Am Benthol Soc 25:730–755.
- Pollard AI, Yuan LL. 2009. Assessing the consistency of response metrics of the invertebrate benthos: a comparison of trait- and identity-based measures. *Freshwater Biol* doi:10.1111/j.1365-2427.2009.02235.x

- Resh VH, Hildrew AG, Statzner B, Townsend CR. 1994. Theoretical habitat templets, species traits, and species richness: a synthesis of long-tern ecological research on the Upper
 Rhône River in the context of concurrently developed ecological theory. *Freshwater Biol* 31:539-554.
- Reynolds CS. 1994. The long, the short and the stalled on the attributes of phytoplankton selected by physical mixing in lakes and rivers. *Hydrobiologia* 289:9-21.
- Reynolds CS, Huszar V, Kruk C, Naselli-Flores L, Melo S. 2002. Towards a functional classification of the freshwater phytoplankton. *J Plankton Res* 24:417-428.
- Reynoldson TB, Logan C, Pascoe T, Thompson S, Sylvestre S. 2001 Invertebrate biomonitoring field and laboratory manual for running water habitats. Canada, Environment Canada, National Waters Research Institute: CABIN: Canadian Aquatic Biomonitoring Network.
- Richards C, Johnson LB, Host GE. 1996. Landscape scale influences on stream habitats and biota. *Can J Fish Aquat Sci* 53:295–311.
- Rubach MN, Ashauer R, Buchwalter DB, De Lange HJ,Hamer M, Preuss TG, Töpke K, Stephen J. Maund SJ. 2010. A framework for trait-based assessment in ecotoxicology. *Integr Environ Assess Manag* this issue.
- Sagnes P, Mérigoux S, Péru N. 2008. Hydraulic habitat use with respect to body size of aquatic insect larvae: Case of six species from a French Mediterranean type stream. *Limnologica* 38(1):23-33.
- Seip KL, Reynolds CS. 1995. Phytoplankton functional attributes along trophic gradient and season. *Limnol Oceanogr* 40:589-597.

- Schmidt-Kloiber A, Graf W, Lorenz A, Moog O. 2006. The AQEM/STAR taxalist a pan-European macro-invertebrate ecological database and taxa inventory. *Hydrobiologia* 566:325–342.
- Shannon CE, Weaver W. 1949. The Mathematical Theory of Communication. Urbana, IL, University of Illinois Press,
- Shipley B, Keddy PA. 1988. The relationship between relative growth rate and sensitivity to nutrient stress in twenty-eight species of emergent macrophytes. *J Ecol* 76:1101-1110.

Simpson JC, Norris RH. 2000. Biological assessment of river quality: development of AUSRIVAS models and outputs. In: Wright JF, Sutcliffe DW, Furse MT, editors.
Assessing the biological quality of fresh waters: RIVPACS and other techniques.
Cumbria, UK, Freshwater Biological Association. p143-163.

Southwood TRE. 1977. Habitat, the templet for ecological strategies? J Anim Ecol 46:337-365.

- Statzner B, Bis B, Dolédec S, Usseglio-polatera P. 2001. Perspectives for biomonitoring at large spatial scales: a unified measure for the functional composition of invertebrate communities in European running waters. *Basic Appl Ecol* 2:73-85.
- Statzner B, Bady P, Dolédec S, Schöll F. 2005. Invertebrate traits for the biomonitoring of large European rivers: an initial assessment of trait patterns in least impacted river reaches. *Freshwater Biol* 50:2136-2161.
- Statzner B, Dolédec S, Hugueny B. 2004. Biological trait composition of European stream invertebrate communities: assessing the effects of various trait filter types. *Ecography* 27:470-488.

- Statzner B, Bonada N, Dolédec S. 2007. Conservation of taxonomic and biological trait diversity of European stream macroinvertebrate communities: a case for a collective public database. *Biodivers Conserv* 16:3609–3632.
- Stevenson RJ, Pan Y, Manoylov KM, Parker CA, Larsen DP, Herlihy AT. 2008. Development of diatom indicators of ecological conditions for streams of the western US. J N Am Benthol Soc 27:1005–1021.
- Tachet H, Richoux P, Bournaud M, Usseglio-Polatera P. 2002. *Invertébrés d'Eau Douce* (2nd corrected impression). CNRS éditions, Paris
- Thiebaut G, Guérold F, Muller S. 2002. Are trophic and diversity indices based on macrophyte communities pertinent tools to monitor water quality? *Water Res* 36:3602–3610.
- Townsend C, Hildrew A. 1994. Species traits in relation to a habitat template for river systems. *Freshw Biol* 31:265-275.
- Tullos DD, Penrose DL, Jennings GD. 2009. Analysis of functional traits in reconfigured channels: implications for the bioassessment and disturbance of river restoration. J N Am Benthol Soc 28:80-92.
- Usseglio-Polatera P, Bournaud M, Richoux P, Tachet H. 2000. Biological and ecological traits of benthic freshwater macroinvertebrates: relationships and definition of groups with similar traits. *Freshwater Biol* 43:175–205.
- Van den Brink PJ, Alexander A, Desrosiers M, Goedkoop W, Goethals P, Liess M, Dyer S.
 2010. Use of traits-based bioassessment approaches in biomonitoring and ecological risk assessment. *Integr Environ Assess Manag* this issue.
- Verberk WCEP, Siepel H, Esselink H. 2008a. Applying life-history strategies for freshwater macroinvertebrates to lentic waters. *Freshwater Biol* 53:1739-1753.

- Verberk WCEP, Siepel H, Esselink H. 2008b. Life-history strategies in freshwater macroinvertebrates. *Freshwater Biol* 53:1722-1738.
- Vieira NKM, Poff NL, Carlisle DM, Moulton SR, Koski ML, Kondratieff BC. 2006. A database of lotic invertebrate traits for North America. U.S. Geological Survey Data Series 187.
 US Geological Survey, US Department of the Interior, Reston, Virginia. (Available from: http://pubs.usgs.gov/ds/ds187/).
- Von der Ohe PC, Liess M. 2004. Relative sensitivity distribution of aquatic invertebrates to organic and metal compounds. *Environ Toxicol Chem* 23:150-156.
- Woodiwiss F. 1960. Trent biotic index of pollution. 2nd Quinquennial Abstract of Statistics relating to the Trent Watershed. Trent River Authority.
- Woodward IF, Cramer W. 2009. Plant functional types and climatic change: an introduction. *J Veg Sci* 7:306-308.
- Wright JF, Furse MT, Armitage PD. 1993. RIVPACS a technique for evaluating the biological quality of rivers in the U. K. *Eur Water Pollut Control* 3:15-25.

Table 1. Selected freshwater examples for algae, aquatic plants, zooplankton, benthic

invertebrates and fish that demonstrate trait-stressor linkages to several common

environmental stressors.

Taxa Group	Stressor	Trait	Trait-Stressor Linkage	Reference
Algae	Nutrients	Cell size	Cell size varies with	Seip and
		trophic status		Reynolds (1995)
	Water	Development	Faster cell division in	Reynolds (1994)
	movement	Rate moving water		
Aquatic	Nutrients	Growth rate	Species with highest	Shipley and
Plants			relative growth rate most sensitive to nutrient stress	Keddy (1988)
	Flood	Vegetative	Type of vegetative	Henry et al.
	disturbance	dispersal	reproduction affects re-	(1996)
			colonization rate	
Zooplankton	Predation	Diapause	Copepods enter diapause to	Hairston and
		stage	avoid fish	Walton (1986)
	Predation	Body size	Zooplankton size decreases	Brooks and
			in presence of predators	Dodson (1965)
Benthic	Deposited	Clinger habit	Decreased abundance with	Pollard and Yuan
Invertebrates	sediments		increased sediment	(2009)
	Channel Mobility and		Higher mobility and more	Tullos et al.
	reconfiguration	development	rapid development with	(2009)
			channel disturbance	
Fish Hydrologic		Body shape	Abundance of deep-bodied	Poff and Allan
	Variability		fish increases with greater	(1995)
			hydrological stability	
	Habitat	Invertivores	Decreased abundance with	Goldstein and
			reduced habitat quality	Meador (2005)

Table 2. Examples of trait databases for freshwater, marine and selected terrestrial habitats summarized for different taxonomic groups. Information for each database is provided for level of taxonomic resolution, method of coding, format in which the information is distributed and a brief description of key aspects of the database. Data not available are indicated by (na).

Habitat	Group	Taxonomic resolution	Coding method	Distribution Format	Description of key aspects of database	Examples
Stream and river	Macroinvertebrates	Variable	Fuzzy	Spreadsheet	European coverage	Tachet et al. (2002); Usseglio-Polatera et al. (2000); Statzner et al. (2007)
	Macroinvertebrates, fish, diatoms	Species	Binary or ten point assignment	Web database	Variable European and taxonomic coverage	Eurolimpacs consortium (2008)
	Macroinvertebrates	Genus	Fuzzy or binary	Spreadsheet	North American lotic insect focus	Vieira et al. (2004); Poff et al. (2006)
	Fish	Species	Binary	Spreadsheet	North American and European coverage limited to six traits	Lamouroux et al. 2002
Freshwater and marine	Phytoplankton	Na	na	na	Defines a structural framework for work in progress	Litchman and Klausmeier (2008)
	Phytoplankton	Genus/ Species	Descriptive data	na	Traits-separated for freshwater functional groups	Reynolds (2002)
	Fish	Species	Descriptive data	Web database	Well-established web- based database	http://www.fishbase.org
Freshwater- terrestrial ecotone	Riparian plants	Na	na	na	Conceptual approach focused on monitoring and management	Merritt et al. (2009)

Terrestrial	Primates	Species	Quantitative variables	Spreadsheet	Global coverage	Cowlishaw et al. (2009)
	Lepidoptera	Species	Binary and quantitative variables	na	Global coverage limited to two families only.	Garcia-Barros (2008)
	Lepidoptera	Species	Quantitative variables	na	Feeding habits of UK butterflies	Hardy et al. (2007)
Multiple habitats	Frogs	Species	Quantitative and qualitative variables	Spreadsheet	Life-history and ecological traits of Australian frog fauna	Murray and Hose (2005a, 2005b)
	Plants	Species	Quantitative variables	Web database	Well-established web- based database	http://www.try-db.org
	Mammals	Species	Quantitative variables	Spreadsheet and web database	Global coverage	http://lynx.ncl.ac.uk/uthe ria/; Jones et al. (2009)

LIST OF FIGURES:

- **Figure 1.** Traditional taxonomic-based bioassessment approaches involve collecting samples from potentially impacted and reference sites, identifying the organisms in those samples, and deriving a metric from the differences in the compositions. Linking traits databases to the taxonomic data allows comparison of reference and impacted sites on the basis of the traits of organisms in those samples. This not only provides additional metrics of the level of impairment, but can also allow diagnosis of impairment cause.
- **Figure 2.** Conceptual diagram that compares the frequency distribution of species traits at a reference and test site. After exposure to a stressor, traits at the test site are modified, or filtered, such that traits that impart resistance or resilience to the stressor are favored. This results in a shift in the frequency distribution of species traits at the test site relative to the reference distribution.



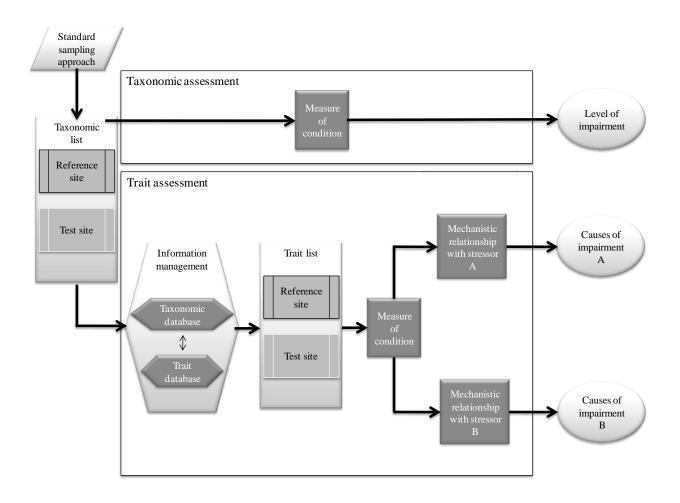


Figure 2.

